Acid-Base Properties of Biological Phenylalkylamines Characterised by CD-pH Titrations

J. Chem. Research (S), 1999, 306–307 J. Chem. Research (M), 1999, 1331–1342

H. Hegedüs, A. Gergely, P. Horváth and B. Noszál*

Semmelweis University, Institute of Pharmaceutical Chemistry H-1092 Budapest, Högyes E. u. 9., Hungary

The acid-base chemistry of eleven selected chiral biological and drug molecules is characterised in terms of log K values by CD-pH titrations.

Characterization of acid-base properties of the biological and xenobiotic phenylalkylamines has been the subject of several studies, using various methods, as surveyed below.

The experimental techniques that can monitor protonation—deprotonation processes in solution include pH-potentiometry, which is certainly the most used method.^{3–5} Of the spectroscopic techniques, UV–VIS photometry is suitable if the basic site is a strong chromophore, such as phenolate or thiolate. However, protonations of less intense chromophores (*e.g.* carboxylate or even more so amines) can only be observed with very limited sensitivity, particularly, when strong chromophores predominate the spectrum. NMR-pH titrations can be quite generally applied when the molecule contains a non-labile proton, or other NMR active nucleus adjacent to the basic site.⁹

When chirally perturbed, protonation-sensitive chromophores exist in a molecule, CD-pH titration may be also an appropriate method to characterise basicities. Provided that molar ellipticity is high, the CD-pH technique can be orders of magnitude more sensitive than pH-potentiometry, throughout the pH scale. Nevertheless, CD-pH titrations have only been sporadically used to determine log *K* values to characterise basicities¹⁷ and resulted in controversial data.¹⁸

In this study the pH-dependent chiroptical properties of eleven biologically active phenylalkylamines were investigated.

The evaluation method was analogous to that used in UV-pH titrations, but exploited the higher CD sensitivity and chromophorial selectivity of these compounds. For the log *K* determination, a minimum of nine solutions of different pH values, and five wavelengths of different ellipticities were used.

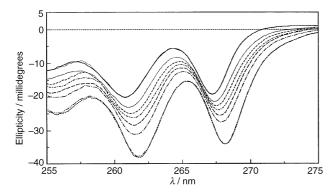


Fig. 3 CD spectra of (-)-selegiline solutions at pH 3.0, 4.0 (top), 7.0, 7.2, 7.45, 7.65, 7.80, and 10.0, 11.0 (bottom). Spectra of the completely protonated (pH 3.0, 4.0) and unprotonated (pH 10.0, 11.0) forms essentially overlap.

Table 2 Protonation constants of chiral phenylalkylamines $(25 \,^{\circ}\text{C}, I = 0.1)$

		Literature	
Compound	р <i>К</i> а ^а	р <i>К</i> _а	Reference
$\overline{(-)-(R)}$ -Selegiline	7.59 ± 0.02	7.48 ± 0.01^{b}	19
(+)- (S) -Selegiline	$\textbf{7.58} \pm \textbf{0.02}$	7.48 ± 0.01^{b}	19
(-)-(R)-Desmethylselegiline	$\textbf{7.83} \pm \textbf{0.06}$	_	_
(-)-(R)-Amphetamine	10.08 ± 0.02	$10.16 \pm 0.06^{b,c}$	20
		10.13 ^{b,d}	21
		10.03	22
		9.77 ± 0.05^{b}	22
		$9.93 \pm 0.01^{b,d}$	23
(-)-(R)-Metamphetamine	10.34 ± 0.05	9.87^{b}	22
(-)- $(1R, 2S)$ -Ephedrine	$\boldsymbol{9.65 \pm 0.07}$	9.58 ± 0.02^{b}	24
. , . , , .		9.56	26
		9.72^{d}	25
		$9.84 \pm 0.02^{b,c}$	27
(+)- $(1S, 2R)$ -Ephedrine	$\boldsymbol{9.70 \pm 0.08}$	9.58 ± 0.02^{b}	24
		$9.84 \pm 0.0^{b,c}$	27
		$9.75 \pm 0.06^{b,c}$	20
(-)- $(1R, 2S)$ -Norephedrine	$\boldsymbol{9.09 \pm 0.06}$	$9.75 \pm 0.06^{b,c}$	20
		$9.75^{b,c}$	30
		9.44 ± 0.04^d	23
(+)- $(1S, 2R)$ -Norephedrine	9.07 ± 0.04	$9.75 \pm 0.06^{b,c}$	20
(,, (,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,		$9.75^{b,c}$	30
$(-)$ - $(1R, 2R)$ - ψ -Ephedrine	$\boldsymbol{9.90 \pm 0.09}$	_	_
$(+)$ - $(1S, 2S)$ - ψ -Ephedrine	9.95 ± 0.05	9.88^{d}	25
(1) (1-,==) y =p 110 d 11110		9.72 ^e	25

^aCD-pH, this work. ^bRacemic. ^cI = 1.0 KNO₃. ^d 20 °C ^e 22 °C.

Basicities of the investigated primary, secondary and tertiary phenylalkylamines reflect several or intramolecular effects, such as N-methylation, C-ethinylation or β -hydroxylation. Basicity differences between ephedrine and ψ -ephedrine could be interpreted in terms of different rotamer populations. Fig. 3 shows pH-dependent spectra of (–)-selegiline, in the aromatic L_B band. Table 2 contains log K values determined in this work, and collected from the literature. Literature data of the same compounds show in some cases discrepancies ≥ 0.3 , that precluded meaningful comparisons, and necessitated a comprehensive study.

Technique used: Ellipsometry

Figures: 4

Tables: 2

References: 32

Received, 11th January 1999; Accepted, 27th January 1999 Paper E/9/00332K

^{*}To receive any correspondence.

References cited in this synopsis

- F. J. C. Rossotti and H. S. Rossotti, The Determination of Stability Constants, McGraw Hill, New York, 1961.
- D. J. Legett, Computational Methods for the Determination of Formation Constants, Plenum Press, New York, London, 1985.
- B. Noszál, Acid-Base Properties of Bioligands, in Biocoordination Chemistry, Coordination Equilibria in Biologically Active Systems, ed. K. Burger, Ellis-Horwood, Chichester, 1990, pp. 18–52.
- D. L. Rabenstein, J. Am. Chem. Soc., 1973, 95, 2797.
- R. Berhallam, E. Collange and M. R. Paris, Bull. Soc. Chim. Fr., 1985, 6, 1159.
- 18
- B. Noszál and M. Erdélyi, manuscript in preparation.K. Takács-Novak and A. Avdeef, J. Pharmaceut. Biomed. 19 Anal., 1996, 14, 1405.
- K. S. Rajan, J. M. Davis, R. W. Colburn and F. H. Jarke, J. Neurochem., 1972, 19, 1099.

- 21 G. P. Lewis, Br. J. Pharmacol., 1954, 9, 488.
- I. D. Chawla and A. C. Andrews, J. Inorg. Nucl. Chem., 1969, **31**, 3809.
- 23 D. H. Everett and J. B. Hyne, J. Chem. Soc., 1958, 1636.
- 24 A. C. Andrews and J. Kirk Romary, J. Chem. Soc., 1964,
- K. S. Rajan, J. M. Davis, R. W. Colburn and F. H. Jarke, J. 25 Neurochem., 1971, 18, 345.
- R. J. Raffa, M. J. Stern and L. Molspeis, Anal. Chem., 1968,
- V. Prelog and O. Häfliger, Helv. Chim. Acta, 1950, 33, 2021.
- K. S. Rajan and J. M. Davis, J. Inorg. Nucl. Chem., 1976, 38,
- 29 G. Girault-Vexlearschi, Bull. Soc. Chim. Fr., 1956, 589.
- E. B. Leffler, H. M. Spencer and A. Burger, J. Am. Chem. Soc., 1951, 73, 2611.